ENVIRONMENTAL CHEMISTRY METHOD EVALUATION REPORT

NUMBER: ECM 0033S1

AN ELISA IMMUNOASSAY METHOD FOR THE DETERMINATION OF RESIDUES OF METHOMYL IN SOIL.

ENVIRONMENTAL CHEMISTRY SECTION (ECS)

ANALYTICAL CHEMISTRY BRANCH

BIOLOGICAL AND ECONOMIC ANALYSIS DIVISION

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PART I

SUMMARY AND CONCLUSION

We have completed an Environmental Chemistry Method Evaluation on methomyl in soil. This method, Dupont number AMR-2396-92, is an enzyme-linked immunoassay (ELISA) for measurement of methomyl over a range of 0.05 to 5.0 parts per billion (ppb).

Following the suggestion of EFGWB, we fortified a soil matrix with methomyl at 5.0, 10, and 50 ppb. In order to evaluate the method at a level near the limit of detection, we also fortified soil at 2.5 ppb. All samples were done in replicates of four at each level. The Dupont method limit of quantitation (LOQ) of 10 ppb was validated by our data. Recoveries for methomyl at the (LOQ) ranged from 116% to 120% with relative standard deviation (RSD) of 1.6%. Although this method involves a simple extraction and no clean-up, we found the recoveries and precision to be good at or above the Limit of Quantitation. The unfortified soil sample analyses showed some methomyl response. This response could be from methomyl already in the soil, or from cross reactivity or matrix effect. Since the amount found in the unfortified soil is less than ten percent of the (LOQ) it is of little consequence.

We feel that the method could be used for low-cost monitoring of soil for methomyl. However, since there can be some cross-reactivity from other compounds, we emphasize the necessity of confirmatory analysis.

We encountered no problems with the method. However, the precoated microplates and some reagents were obtained from the registrant since they are not readily available from commercial sources. EFED should make sure that Dupont understands that they are responsible for making the precoated microplates commercially available or for licensing their technology to one of the kit manufacturers. They are responsible for making the test kits commercially available to all potential users.

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PART II

ANALYTICAL RESULTS FOR METHOMYL

EPA RECOVERIES IN SOIL

Sample no.	Added (ppb)	Found (ppb)	Recovery Data		
01	0	1.00	mean(ppb) = 0.75		
02	0 .	0.62	sd = .226		
03	0	0.50	rsd = 30.2%		
04	0	0.86	•		
		2 54	maan(nnh) = 2 65		
26	2.50	3.54	mean(ppb) = 3.65 sd = .395		
27	2.50	3.32			
28	2.50	3.50	rsd = 10.8%		
29	2.50	4.22	mean recovery= 146%		
05	5.00	6.18	mean(ppb) = 6.52		
06	5.00	6.60	sd = .252		
07	5.00	6.78	rsd = 3.9%		
08	. 5.00	6.50	mean recovery = 130%		
09	10.0	11.8	mean(ppb) = 11.7		
10	10.0	11.6	sd = .192		
11	10.0	12.0	rsd = 1.6%		
12	10.0	11.6	mean recovery = 117%		
13	50.0	49.4	mean(ppb) = 49.0		
14	50.0	45.8	sd = 2.63		
15	50.0	48.6	rsd = 5.4%		
16	50.0	52.2	mean recovery = 98%		

PART III

EXPERIMENTAL SUMMARY

(a) Principle of Method

Polyclonal anti-methomyl antibodies(Ab) and buffer are added to a sample containing an unknown amount of methomyl and incubated. The antibodies bind to methomyl molecules present in the sample (if any).

Aliquots of the solution are added to wells on a 96-well microplate, which have been coated with a methomyl derivative-Ovalbumin conjugate. Any excess (Ab) not bound to methomyl in the sample will bind to methomyl immobilized on the microwell plate. The plate is then washed to remove any (Ab) not bound to the plate. The amount of (Ab) bound to the microwell is an inverse measure of the amount of methomyl in the sample.

To detect the (Ab) bound to each microwell, an anti-rabbit antibody conjugated to an enzyme, alkaline phosphatase(Ab-E), is added to each well and incubated. This (Ab-E) will bind to any anti-methomyl antibodies bound to the microwell. The microwell plate is then washed to remove any unbound (Ab-E).

The alkaline phosphatase substrate, para-nitrophenyl-phosphate, is added to the microwells. The enzyme-substrate reaction produces a yellow color which is inversely proportional to the concentration of methomyl in the sample. The Microplate Reader quantitates absorbance in each well at 405 nm. Computer software is used to construct a standard curve from standards run on the plate, and to calculate the methomyl concentrations of each unknown on the plate.

(b) Source of Analytical Reference Standard

Methomyl standard was supplied and certified at 99.77% by E.I.Dupont de Nemours & company, Wilmington, Del.

(c) Source of Sample Matrix

Soil was obtained from Dupont from a supply of soils used in their recovery trials. The soil was characterized as silt loam with 2.3% organic matter and pH of 6.6.

(d) <u>Instrumentation for Quantitation</u>

- 1. Vmax Kinetic Microplate Reader with 405 nm optical filter. Molecular Devices Corporation.
- ULTRAWASH PLUS, automatic microplate washer/aspirator, Dynatech Laboratories, Inc.

(e) Modification of Method

We used microplates supplied by Dupont and already coated with Coating Antigen Reagent. The method includes a procedure for coating blank microplates.

(f) Sample Extraction

A 10 gram soil sample is extracted with 20 ml of acetone/phosphate buffered saline solution (PBS). The supernatant is filtered and a 1.0 ml aliquot is brought to near dryness under nitrogen and reconstituted with 10 ml of phosphate buffered saline solution (PBS).

(g) Calculations

The absorbance of each of the 96 wells of the microwell plate is read on a microplate reader equipped with a 405 nm filter and processed by a computer program which generates a standard curve based upon a four-parameter logit function. The standard four-parameter logit curve generated is sigmoidal in shape with optical density (OD) on the Y-axis and the log of the concentration of methomyl on the X-axis. Each calibration standard and sample is pipetted into three wells of the microplate and analyzed in triplicate. The average of the optical density (OD) readings for these three wells is then used to interact with the standard curve to calculate the concentration of methomyl in each sample. For comparison, a semi-log curve fit would provide reasonably good data and can be used to check the calculations.

(h) Graphs and Data

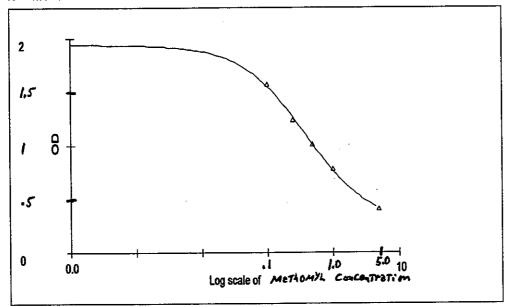
The following pages contain a print-out of the standard curve and data generated by the methomyl calibration standards and selected samples.

Curve Fit: 4-Parameter

Corr. Coeff: 1.00

 $y = (A-D)/(1 + (x/C)^AB) + D$

A= 1.95 B= 0.890 C= 0.391 D= 0.265



STANDARD STD01	Std. Value 0	Well 82 C2 D2	OD 1.964 1.891 1.974	Mean 1.943	Std Dev 0.045	CV 2.332	Calc. Value <<<< 0.009 <<<<	Sample ID
STD02	0.1	B3 C3 D3	1.588 1.561 1.583	1.577	0.014	0.911	0.090 0.100 0.092	
STD03	0.25	B4 C4 D4	1.295 1.247 1.201	1.247	0.047	3.769	0.234 0.267 0.303	
STD04	0.5	85 C5 D5	1.050 0.997 1.004	1.017	0.029	2.832	0.454 0.523 0.514	
STD05	1	B6 C6 D6	0.808 0.767 0.777	0.784	0.021	2.728	0.898 1.020 0.989	
STD06	5	B7 C7 D7	0.428 0.399 0.429	0.418	0.017	4.074	4.796 6.106 4.760	

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DATA FILE: DESCRIPTION: PROTOCOL: DESCRIPTION:

11169405

methomyl

MODE: WAVELENGTH: CALIBRATION: Endpoint 405 On AUTOMIX:

On

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						<u> </u>	
	PLATE BLANK BL	Mean OD 0.108	Std Dev 0.002	CV 1.580	Well A2 A3 A4 A5 A6 A7 A8 A9 A10	OD 0.108 0.107 0.106 0.108 0.109 0.109 0.108 0.110 0.112 0.107	Sample ID SUBSTRATE BLANKS
	STANDARDS STD01	Mean OD 1.943	Std Dev 0.045	CV 2.332	Well B2 C2 D2	OD 1.964 1.891 1.974	Sample ID 0.0 mg/ml 0.1 mg/ml 0.25 mg/ml 0.50 mg/ml 1.0 mg/ml
	STD02	1.577	0.014	0.911	83 C3 D3	1.588 1.561 1.583	0.1 ns/ml
	STD03	1.247	0.047	3.769	B4 C4 D4	1.295 1.247 1.201	0.25 ns/ml
•	STD04	1.017	0.029	2.832	B5 C5 D5	1.050 0.997 1.004	0.50 Mafal
	STD05	0.784	0.021	2.728	86 C6 D6	0.808 0.767 0.777	1.0 ms/ml
	STD06	0.418	0.017	4.074	B7 C7 D7	0.428 0.399 0.429	5.0 ms/ml
-	UNKNOWNS UNK01	Mean OD 1.719	Std Dev 0.080	CV 4.672	Well B8 C8 D8	OD 1.803 1.711 1.643	Factor PPb Sample ID 20.00 0.995 Unfortified Sail
•	UNK02	1.794	0.100	5.579	B9 C9 D9	1.874 1.827 1.682	20.00 0.619
	UNK03	1.816	0.061	3.336	B10 C10 D10	1.879 1.813 1.758	20.00 0.493
	UNK04	1.740	0.022	1.249	B11 C11 D11	1.749 1.757 1.716	20.00 0.855

MOLECULAR DEVICES

UNKNOWNS UNK09	Mean OD 0.954	Std Dev 0,010	CV 1.091	Well E6 F6 G6	OD 0.966 0.946 0.951	.Factor 20.00	<u>PPL</u> 11.77	Sample ID Fortifical AT 10 PPb
UNK10	0.959	0.034	3.558	E7 F7 G7	0.981 0.920 0.977	20.00	11.65	<i>1</i> ′
UNK11	0.947	0.014	1.510	E8 F8 G8	0.963 0.935 0.944	20.00	12.00	,,
UNK12	0.961	0.017	1.770	E9 F9 G9	0.968 0.942 0.974	20.00	11.55	+ f
UNK13	0.538	0.010	1.881	E10 F10 G10	0.550 0.533 0.532	20.00	49.37	FORTIFIEL BT 50 175
UNK14	0.554	0.019	3.340	E11 F11 G11	0.564 0.566 0.533	20.00	45.85	
UNK15	0.541	0.006	1.174	H2 H3 H4	0.545 0.545 0.534	20.00	48.61	, ,
UNK16	0,528	0.024	4.618	H5 H6 H7	0.519 0.556 0.510	20.00	52.23	11